#### Better than Chance: the importance of null models

Kevin Karplus

karplus@soe.ucsc.edu

Biomolecular Engineering Department
Undergraduate and Graduate Director, Bioinformatics
University of California, Santa Cruz



#### **Outline of Talk**

- What is a null model (or null hypothesis) for?
- Example 1: is a conserved ORF a protein?
- Example 2: is residue-residue contact prediction better than chance?
- Example 3: how should we remove composition biases in HMM searches?



## Scoring (Bayesian view)

- Model M is a computable function that assigns a probability  $P(A \mid M)$  to each sequence A.
- When given a sequence A, we want to know how likely the model is. That is, we want to compute something like  $P(M \mid A)$ .
- Bayes Rule:

$$P(M \mid A) = P(A \mid M) \frac{P(M)}{P(A)}$$
.

 $\triangleleft$  Problem: P(A) and P(M) are inherently unknowable.



#### **Null models**

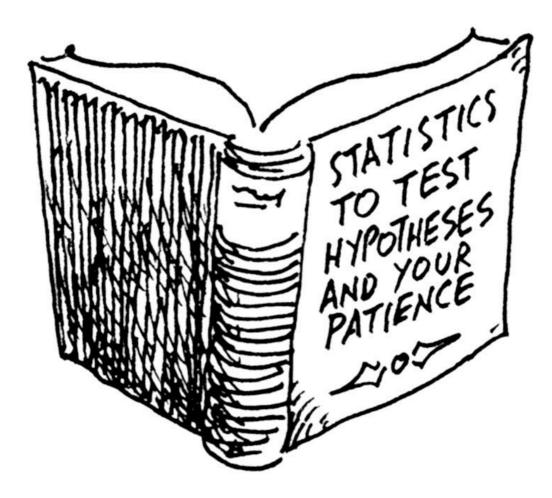
Standard solution: ask how much more likely M is than some null hypothesis (represented by a null model):

$$\frac{\mathsf{P}(M \mid A)}{\mathsf{P}(N \mid A)} = \frac{\mathsf{P}(A \mid M)}{\mathsf{P}(A \mid N)} \qquad \frac{\mathsf{P}(M)}{\mathsf{P}(N)} .$$

posterior odds likelihood ratio prior odds



### Test your hypothesis



Thanks to Larry Gonick The Cartoon Guide to Statistics



## Scoring (frequentist view)

- We believe in models when they give a large score to our observed data.
- Statistical tests (p-values or E-values) quantify how often we should expect to see such good scores "by chance".
- These tests are based on a null model or null hypothesis.



### Small p-value to reject null hypothesis



Thanks to Larry Gonick The Cartoon Guide to Statistics



## Statistical Significance (2 approaches)

Markov's inequality For any scoring scheme that uses

$$\ln \frac{\mathsf{P}\left(\mathsf{seq} \mid M\right)}{\mathsf{P}\left(\mathsf{seq} \mid N\right)}$$

the probability of a score better than T is less than  $e^{-T}$  for sequences distributed according to N.

Parameter fitting For "random" sequences drawn from some distribution other than N, we can fit a parameterized family of distributions to scores from a random sample, then compute P and E values.



#### **Null models**

- P-values (and E-values) often tell us nothing about how good our hypothesis is.
- What they tell us is how bad our null model (null hypothesis) is at explaining the data.
- A badly chosen null model can make a very wrong hypothesis look good.



## **Example 1: long ORF**

- A colleague found an ORF in an archæal genome that was 388 codons long and was wondering if it coded for a protein and what the protein's structure was.
- We know that short ORFs can appear "by chance".
- So how likely is this ORF to be a chance event?



#### **Null Model 1**

- DNA is undergoing no selection at all
- ← G+C content bias. (GC is 36.7%, AT is 63.3%.)
- Probability of stop codon TAG= 0.3165\*0.3165\*0.1835=0.0184, TGA=0.0184, TAA=0.0317, so p(STOP)=0.0685.
- **4** P(388 codons without stop) =  $(1 p(STOP))^{388}$  = 1.1e-12
- E-value in a 3 Megabase genome is about 3.3e-6.
- We can easily reject the null hypothesis!



#### Null Model 2

- I forgot to tell you: this ORF is on the opposite strand of a known 560-codon ribosomal gene.
- What is the probability of this long an ORF, on opposite strand of known gene?
- Generative model: simulate random codons using the codon bias of the organism, take reverse complement, and see how often ORFs 388-long or longer appear.
- Taking 100,000 samples, we get estimates of P-value in the range 3e-05 to 6e-05.
- There are about 3000 genes, giving us an E-value of 0.09 to 0.18.

#### **Null Model 3**

- We can do the same sort of simulation, but restrict the codons to ones that would code for exactly the same protein on the forward strand.
- Now we get P-value of around 0.01 for long ORFs on the reverse strand of genes coding for this protein.



#### **Protein or chance ORF?**



Thanks to Larry Gonick The Cartoon Guide to Statistics



### Not a protein

- A tblastn search with the sequence revealed similar ORFs in many genomes.
- All are on opposite strand of homologs of same gene.
- "Homologs" found by tblastn often include stop codons.
- There is no evidence for a TATA box upstream of the ORF.
- No strong evidence for selection beyond that explained by known gene.

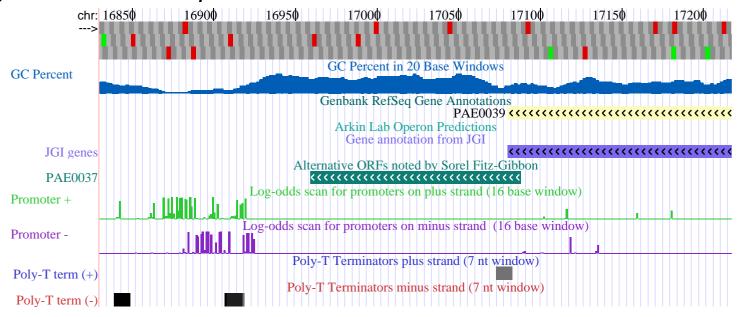
Conclusion: it is rather unlikely that this ORF encodes a protein.



### Example 1b: another ORF

pae0037: ORF, but probably not protein gene in

Pyrobaculum aerophilum



- Promoter on wrong side of ORF.
- High GC content (need local, not global, null)
  - Strong RNA secondary structure.

#### **Example 2: contacts**

- Is residue-residue contact prediction better than chance?
- Early predictors (1994) reported results that were 1.4 to5.1 times "better than chance" on a sample of 11 proteins.
- But they used a uniform null model:

P(residue i contacts residue j) = constant .

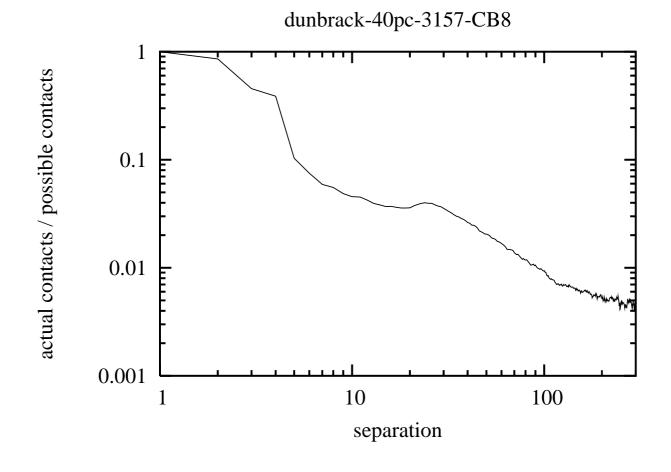
A better null model:

P (residue 
$$i$$
 contacts residue  $j$ ) = P (contact | separation =  $|i-j|$ ) .



### P(contact|separation)

Using CASP definition of contact, CB within 8 Å, CA for GLY.





## Can get accuracy of 100%

- By ignoring chain separations, the early predictors got what sounded like good accuracy (0.37–0.68 for L/5 predicted contacts)
- But just predicting that i and i+1 are in contact would have gotten accuracy of 1.0 for even more predictions.
- More recent work has excluded small-separation pairs, with different authors choosing different thresholds.
- $\leq$  CASP uses separation  $\geq 6$ ,  $\geq 12$ , and  $\geq 24$ , with most focus on > 24.



## **Evaluating contact prediction**

Two measures of contact prediction:

Accuracy:

$$\frac{\sum \chi(i,j)}{\sum 1}$$

Weighted accuracy:

$$\frac{\sum \frac{\chi(i,j)}{\mathsf{P}\big(\mathsf{contact}|\mathsf{separation}=|i-j|\big)}}{\sum 1}$$

= 1 if predictions no better than chance, independent of separations for predicted pairs.



### Separation as predictor

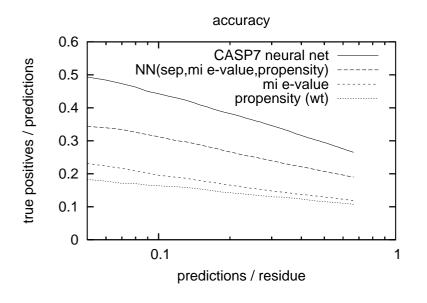
If we predict all pairs with given separation as in contact, we do much better than uniform model.

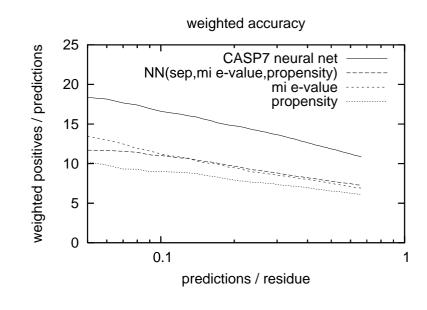
sep	$P\left(contact\ \Big \  i-j =sep ight)$	$P\left(contact \;\middle \;  i-j  \geq sep ight)$	"better than chance"
6	0.0751	0.0147	4.96
9	0.0486	0.0142	3.42
12	0.0424	0.0136	3.13
24	0.0400	0.0116	3.46



## Now doing better

separation  $\geq 9$  contacts/residue taken separately for each protein

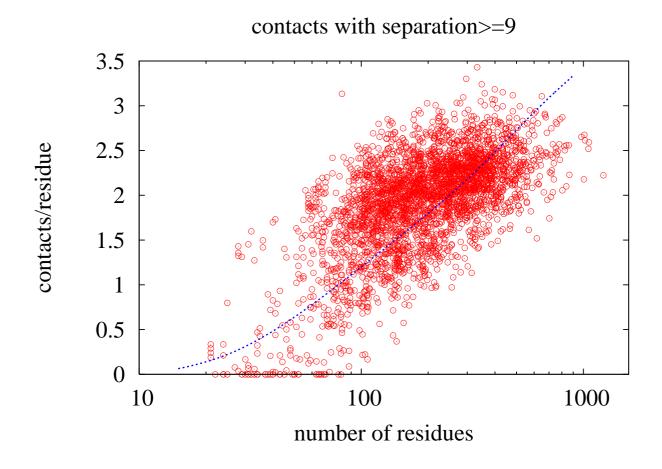






## Contacts per residue

We can also use our null model to predict the number of contacts per residue (which is not a constant).





## Example 3: HMM

- Hidden Markov models assign a probability to each sequence in a protein family.
- A common task is to choose which of several protein families (represented by different HMMs) a protein belongs to.



#### **Standard Null Model**

Null model is an i.i.d (independent, identically distributed) model.

$$\mathsf{P}\left(A \mid N, \mathsf{len}\left(A\right)\right) = \prod_{i=1}^{\mathsf{len}(A)} \mathsf{P}(A_i) \; .$$

$$\mathsf{P}\left(A \mid N\right) = \mathsf{P}(\text{sequence of length len}\left(A\right))$$
 
$$\prod_{i=1}^{\mathsf{len}(A)} \mathsf{P}(A_i) \; .$$



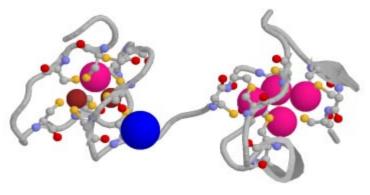
### Composition as source of error

- When using the standard null model, certain sequences and HMMs have anomalous behavior. Many of the problems are due to unusual composition—a large number of some usually rare amino acid.
- For example, metallothionein, with 24 cysteines in only 61 total amino acids, scores well on any model with multiple highly conserved cysteines.

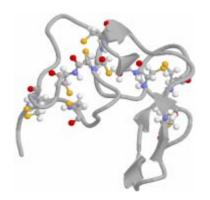


## **Composition examples**

Metallothionein Isoform II (4mt2)



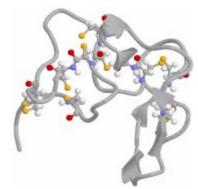
Kistrin (1kst)



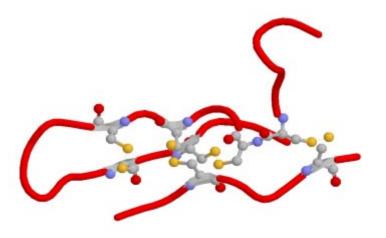


### **Composition examples**

Kistrin (1kst)



Trypsin-binding domain of Bowman-Birk Inhibitor (1tabl)





#### Reversed model for null

- We avoid this (and several other problems) by using a reversed model  $M^r$  as the null model.
- The probability of a sequence in  $M^r$  is exactly the same as the probability of the reversal of the sequence given M.
- This method corrects for composition biases, length biases, and several subtler biases.



# Helix examples

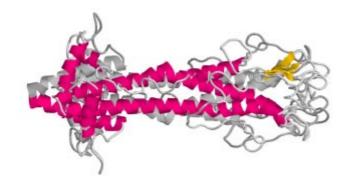
Tropomyosin (2tmaA)



Colicin la (1cii)

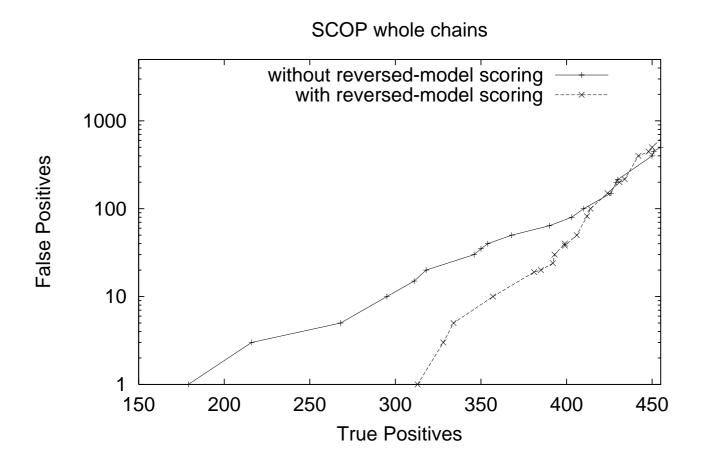


Flavodoxin mutant (1vsgA)





### Improvement from reversed model





### Take-home messages

- Base your null models on biologically meaningful null hypotheses, not just computationally convenient math.
- Generative models and simulation can be useful for more complicated models.
- Picking the right model remains more art than science.



#### Web sites

**List of my papers:** http://www.soe.ucsc.edu/~karplus/papers/

These slides: http://www.soe.ucsc.edu/~karplus/papers/

better-than-chance-jul-07.pdf

Reverse-sequence null: Calibrating E-values for hidden Markov models with

reverse-sequence null models. *Bioinformatics*, 2005. 21(22):4107–4115;

doi:10.1093/bioinformatics/bti629

Archæal genome browser: http://archaea.ucsc.edu

**UCSC** bioinformatics (research and degree programs) info:

http://www.soe.ucsc.edu/research/compbio/

