Bayesian Modeling, Inference, Prediction and Decision-Making

5: Bayesian Model Specification (Section 2)

David Draper

Department of Applied Mathematics and Statistics University of California, Santa Cruz

> draper@ams.ucsc.edu www.ams.ucsc.edu/~draper

> > eBay/Google

10 Fridays, 11 Jan-22 Mar 2013 (except 25 Jan)

Short course web page:

www.ams.ucsc.edu/~draper/eBay-Google-2013.html

(This section of the notes is based in part on unpublished joint work with a recent Ph.D. student of mine, Milovan Krnjajić.)

© 2013 David Draper (all rights reserved)

What is a Bayesian Model?

Definition: A **Bayesian model** is a **mathematical framework** (embodying **assumptions** and **judgments**) for **quantifying uncertainty about unknown quantities** by relating them to **known quantities**.

Desirable for the **assumptions** and **judgments** in the model to arise as directly as possible from **contextual information** in the problem under study.

The most satisfying approach to achieving this goal appears to be that of de Finetti (1990): a Bayesian model is a joint predictive distribution

$$p(y) = p(y_1, \dots, y_n) \tag{1}$$

for as-yet-unobserved **observables** $y = (y_1, \ldots, y_n)$.

Example 1: Data = health outcomes for all patients at one hospital with heart attack admission diagnosis.

Simplest possible: $y_i = 1$ if patient *i* dies within 30 days of admission, 0 otherwise.

de Finetti (1930): in absence of any other information, my predictive uncertainty about y_i is exchangeable.

Representation theorem for binary data: if I'm willing to regard (y_1, \ldots, y_n) as part of an **infinitely exchangeable sequence** (meaning that I judge all **finite subsets** exchangeable; this is like **thinking** of the y_i as having been **randomly sampled** from the **population** (y_1, y_2, \ldots)), then to be **coherent** my joint predictive distribution $p(y_1, \ldots, y_n)$ must have the simple **hierarchical** form

$$\begin{array}{l} \theta \sim p(\theta) \\ (y_i|\theta) \stackrel{\text{IID}}{\sim} \quad \text{Bernoulli}(\theta), \end{array}$$
(2)

where $\theta = P(y_i = 1) =$ limiting value of mean of y_i in infinite sequence.

Model = Prior (Sometimes)

Mathematically $p(\theta)$ is mixing distribution in

$$p(y_1,\ldots,y_n) = \int_0^1 \theta^s (1-\theta)^{n-s} p(\theta) \, d\theta, \qquad (3)$$

where $s = \sum_{i=1}^{n} y_i$; statistically, $p(\theta)$ provides opportunity to quantify prior information about θ and combine with information in y.

Thus, in simplest situation, **Bayesian model** specification = choice of scientifically appropriate prior distribution $p(\theta)$.

Example 2 (elaborating Example 1): Now I want to predict real-valued **sickness-at-admission score** instead of mortality (still no **covariates**).

Uncertainty about y_i still exchangeable; de Finetti's (1937) representation theorem for real-valued data: if (y_1, \ldots, y_n) part of infinitely exchangeable sequence, all coherent joint predictive distributions $p(y_1, \ldots, y_n)$ must have hierarchical form

$$\begin{array}{ccc} F & \sim & p(F) \\ (y_i|F) & \stackrel{\text{IID}}{\sim} & F, \end{array}$$
 (4)

where F = limiting empirical cumulative distribution function (CDF) of infinite sequence $(y_1, y_2, ...)$.

Bayesian Nonparametrics

Thus here Bayesian model specification = choosing scientifically appropriate mixing (prior) distribution p(F) for F.

However, F is **infinite-dimensional parameter**; putting probability distribution on $\mathcal{D} = \{all \text{ possible CDFs}\}$ is harder.

Specifying distributions on **function spaces** is task of Bayesian **nonparametric** (BNP) modeling (e.g., Dey et al. 1998).

Example 3 (elaborating Example 2): In practice, in addition to **outcomes** y_i , **covariates** x_{ij} will typically be available.

For instance (Hendriksen et al. 1984), 572 elderly people randomized, 287 to control (*C*) group (standard care) and 285 to treatment (*T*) group (standard care plus in-home geriatric assessment (IHGA): preventive medicine in which each person's medical/social needs assessed, acted upon individually).

One important **outcome** was **number of hospitalizations** (in two years).

 y_i^T , y_j^C = numbers of hospitalizations for **treatment** person *i*, **control** person *j*, respectively.

Suppose treatment/control (T/C) status is only available covariate.

Conditional Exchangeability

Unconditional judgment of exchangeability across all 572 outcomes no longer automatically scientifically appropriate.

Instead **design of experiment** compels (at least initially) judgment of **conditional exchangeability given T/C status** (e.g., de Finetti 1938, Draper et al. 1993), as in

 $(F_T, F_C) \sim p(F_T, F_C) (5)$ $(y_i^T | F_T, F_C) \sim F_T | (y_j^C | F_T, F_C) \sim F_C$

This framework, in which (a) covariates specify conditional exchangeability judgments, (b) de Finetti's representation theorem reduces model specification task to placing appropriate prior distributions on CDFs, covers much of field of statistical inference/prediction.

Note that even in this **rather general nonparametric framework** it will be necessary to have a **good tool** for **discriminating between the quality of two models** (here: **unconditional** exchangeability ($F_T = F_C$; T has **same effect** as C) versus **conditional** exchangeability ($F_T \neq F_C$; T and C effects **differ**)).

Data-Analytic Model Specification

Basic problem of Bayesian model choice: Given future observables $y = (y_1, \ldots, y_n)$, I'm uncertain about y (first-order), but I'm also uncertain about how to specify my uncertainty about y (second-order).

Standard (data-analytic) approach to model specification involves initial choice, for structure of model, of standard parametric family, followed by modification of initial choice—once data begin to arrive—if data suggest deficiencies in original specification.

This approach (e.g., Draper 1995) is **incoherent** (unless I pay an **appropriate price** for **shopping around** for the model): it uses data both to specify **prior distribution on structure space** and to **update** using **data-determined prior** (result will typically be **uncalibrated** (too narrow) predictive distributions for future data).

Dilemma is example of **Cromwell's Rule** (if $p(\theta) = 0$ then $p(\theta|y) = 0$ for all y): initial model choice placed **0 prior probability** on **large regions of model space**; formally all such regions **must also have 0 posterior probability** even if data indicate **different prior on model space** would have been better.

Two Possible Solutions

If use prior on F that places non-zero probability on all Kullback-Leibler neighborhoods of all densities (Walker et al. 2003; e.g., Pólya trees, Dirichlet process mixture priors, when chosen well), then BNP directly avoids Cromwell's Rule dilemma, at least for large n: as n → ∞ posterior on F will shrug off any incorrect details of prior specification, will fully adapt to actual data-generating F (NB this assumes correct exchangeability judgments).

• **Three-way cross-validation** (3CV; Draper and Krnjajić 2005): taking usual cross-validation idea one step further,

(1) Partition data at random into *three* (non-overlapping and exhaustive) subsets S_i .

(2) Fit tentative {likelihood + prior} to S_1 . Expand initial model in all feasible ways suggested by data exploration using S_1 . Iterate until you're happy.

(3) Use final model (fit to S_1) from (2) to create predictive distributions for all data points in S_2 . Compare actual outcomes with these distributions, checking for **predictive calibration**. Go back to (2), change likelihood as necessary, **retune prior** as necessary, to get good calibration. **Iterate** until you're happy.

(4) Announce final model (fit to $S_1 \cup S_2$) from (3), and report **predictive calibration** of this model on data points in S_3 as indication of how well it would perform with new data.

With large *n* probably only need to do this **once**; with **small** and **moderate** *n* probably best to **repeat** (1–4) several times and **combine** results in some appropriate way (e.g., **model averaging**).

Model Selection as a Decision Problem

Given method like 3CV which permits hunting around in model space without forfeiting calibration, two kinds of model specification questions (in both parametric and nonparametric Bayesian modeling) arise:

(1) Is M_1 better than M_2 ? (this tells me when it's OK to discard a model in my search)

(2) Is M_1 good enough? (this tells me when it's OK to stop searching)

It would seem self-evident that to specify a model you have to say to what purpose the model will be put, for how else can you answer these two questions?

Specifying this purpose demands **decision-theoretic basis for model choice** (e.g., Draper 1996; Key et al. 1998).

To take two examples,

(Case 1) If you're going to choose which of several ways to behave in future, then model has to be good enough to reliably aid in choosing best behavior (see, e.g., Draper and Fouskakis example below); or

(Case 2) If you wish to make scientific summary of what's known, then—remembering that hallmark of good science is good prediction—the model has to be good enough to make sufficiently accurate predictions of observable outcomes (in which dimensions along which accuracy is to be monitored are driven by what's scientifically relevant; see, e.g., log score results below).

Utility-Based Variable Selection

Example 4 (Case 1): Draper and Fouskakis (2000, 2004) (also see Fouskakis and Draper 2002) give one example of decision-theoretic model choice in action, demonstrating that **variable selection in regression models** should often be governed by principle that final model should only contain variables that predict well enough **given how much they** cost to collect (see the figure below, which compares $2^{14} = 16,384$ models).



Estimated expected utility as function of number of predictor variables, in problem involving construction of cost-effective scale to measure sickness at hospital admission of elderly pneumonia patients. Best models only have 4–6 sickness indicators out of 14 possible predictors.

Choosing Utility Function

Any reasonable utility function in Example 4 will have two components, one quantifying **data collection costs** associated with construction of given sickness scale, other rewarding and penalizing scale's **predictive successes, failures**.

(Case 2) Sometimes the main goal instead is summary of
 scientific knowledge, which suggests (as noted above) a
 utility function that rewards predictive accuracy.

How can such a **utility function** be specified in a **reasonably general way** to answer **model specification question (1)** above? (Is M_1 better than M_2 ?)

Need scoring rule that measures discrepancy between observation y^* and predictive distribution $p(\cdot|y, M_i)$ for y^* under model M_i given data y.

As noted (e.g.) by Good (1950) and O'Hagan and Forster (2004), **the optimal (impartial, symmetric, proper)** scoring rules are linear functions of $\log p(y^*|y)$.

On calibration grounds it would seem to be a mistake to use data twice in measuring this sort of thing (once to make predictions, again with same data to see how good they are; but ...).

Out-of-sample predictive validation (e.g., Geisser and Eddy 1979, Gelfand et al. 1992) solves this problem: e.g., successively remove each observation y_j one at a time, construct predictive distribution for y_j based on y_{-j} (data vector with y_j removed), see where y_j falls in this distribution.

Log Score as Utility

This motivates **cross-validated** version of **log scoring rule** (e.g., Gelfand and Dey 1994; Bernardo and Smith 1994): with n data values y_j , when choosing among k models $M_i, i \in I$, find that model M_i which maximizes

$$LS_{CV}(M_i|y) = \frac{1}{n} \sum_{j=1}^{n} \log p(y_j|M_i, y_{-j}).$$
(6)

It has been argued that this can be given direct decision-theoretic justification: with utility function for model i

$$U(M_i|y) = \log p(y^*|M_i, y),$$
 (7)

where y^* is **future data value**, expectation in MEU is over **uncertainty about** y^* ; Gelfand et al. (1992) and Bernardo and Smith (1994) claim that this expectation can be accurately **estimated** (assuming exchangeability) by (6) (I'll revisit this claim below).

With approximately **Gaussian** predictive distributions it can also be revealing to compute **predictive** z-scores, for observation j under model i:

$$z_{ij} = \frac{y_j - E(y_j | M_i, y_{-j})}{\sqrt{V(y_j | M_i, y_{-j})}}.$$
(8)

For good predictive calibration of M_i , $\{z_{ij}, j = 1, ..., n\}$ should have mean 0, standard deviation (SD) 1; often find instead that SD is larger than 1 (predictive uncertainty bands not wide enough).

Approximating Log Score Utility

With large data sets, in situations in which predictive distribution has to be estimated by MCMC, direct calculation of LS_{CV} is computationally expensive; need fast approximation to it.

To see how this might be obtained, examine log score in simplest possible model M_0 : for i = 1, ..., n,

$$\mu \sim N(\mu_0, \sigma_\mu^2)$$

$$(Y_i|\mu) \stackrel{\text{IID}}{\sim} N(\mu, \sigma^2)$$
(9)

with σ known, take **highly diffuse prior** on μ so that **posterior** for μ is approximately

$$(\mu|y) = (\mu|\bar{y}) \stackrel{\cdot}{\sim} N\left(\bar{y}, \frac{\sigma^2}{n}\right), \tag{10}$$

where $y = (y_1, ..., y_n)$.

Then **predictive distribution** for next observation is approximately

$$(y_{n+1}|y) = (y_{n+1}|\bar{y}) \stackrel{\cdot}{\sim} N\left[\bar{y}, \sigma^2\left(1+\frac{1}{n}\right)\right], \qquad (11)$$

and LS_{CV} , ignoring linear scaling constants, is

$$LS_{CV}(M_0|y) = \sum_{j=1}^n \ln p(y_j|y_{-j}), \qquad (12)$$

where as before y_{-j} is y with observation j set aside.

But by same reasoning

$$p(y_j|y_{-j}) \doteq N(\bar{y}_{-j}, \sigma_n^2), \qquad (13)$$

where \bar{y}_{-j} is sample mean with observation j omitted,

$$\sigma_n^2 = \sigma^2 \left(1 + \frac{1}{n-1} \right)$$
, so that

$$\ln p(y_j|y_{-j}) \doteq c - \frac{1}{2\sigma_n^2} (y_j - \bar{y}_{-j})^2 \text{ and}$$
$$LS_{CV}(M_0|y) \doteq c_1 - c_2 \sum_{j=1}^n (y_j - \bar{y}_{-j})^2 \quad (14)$$

for some **constants** c_1 and c_2 with $c_2 > 0$.

Now it's **interesting fact** (related to behavior of **jackknife**), which you can prove by **induction**, that

$$\sum_{j=1}^{n} (y_j - \bar{y}_{-j})^2 = c \sum_{j=1}^{n} (y_j - \bar{y})^2$$
(15)

for some c > 0, so finally for $c_2 > 0$ the **result** is that

$$LS_{CV}(M_0|y) \doteq c_1 - c_2 \sum_{j=1}^n (y_j - \bar{y})^2,$$
 (16)

i.e., in this model log score is almost perfectly negatively correlated with sample variance.

But in this model the **deviance** (minus twice the log likelihood) is

$$D(\mu) = -2 \ln l(\mu|y) = c_0 - 2 \ln p(y|\mu)$$

= $c_0 + c_3 \sum_{j=1}^n (y_j - \mu)^2$ (17)

for some $c_3 > 0$, encouraging suspicion that log score should be strongly related to deviance.

Deviance Information Criterion (*DIC***)**

Given parametric model $p(y|\theta)$, Spiegelhalter et al. (2002) define **deviance information criterion** (*DIC*) (by analogy with other information criteria) to be estimate $D(\bar{\theta})$ of model (lack of) **fit** (as measured by deviance) plus **penalty for complexity** equal to twice **effective number of parameters** p_D of model:

$$DIC(M|y) = D(\bar{\theta}) + 2\,\hat{p}_D,\tag{18}$$

where $\overline{\theta}$ is posterior mean of θ ; they suggest that models with **low** *DIC* value are to be **preferred** over those with higher value.

When p_D is difficult to read directly from model (e.g., in complex hierarchical models, especially those with random effects), they motivate the following estimate, which is easy to compute from standard MCMC output:

$$\widehat{p}_D = \overline{D(\theta)} - D(\overline{\theta}), \qquad (19)$$

i.e., difference between **posterior mean of deviance** and **deviance evaluated at posterior mean** of parameters (WinBUGS release 1.4 will **estimate** these quantities).

In model M_0 , p_D is of course 1, and $\bar{\theta} = \bar{y}$, so

$$DIC(M_0|y) = c_0 + c_3 \sum_{j=1}^n (y_j - \bar{y})^2 + 2$$
 (20)

and conclusion is that

$$-DIC(M_0|y) \doteq c_1 + c_2 LS_{CV}(M_0|y)$$
(21)

for $c_2 > 0$, i.e., (if this generalizes) choosing model by maximizing LS_{CV} and by minimizing DIC are approximately equivalent behaviors.

(This connection was **hinted at** in discussion of Spiegelhalter et al. 2002 but never really made **explicit**.)

$LS_{CV} \leftrightarrow DIC$?

Milovan and I are now (work in progress) exploring the scope of (21); in several simple models M so far we find for $c_2 > 0$ that

$$-DIC(M|y) \doteq c_1 + c_2 LS_{CV}(M|y), \qquad (22)$$

i.e., across repeated data sets generated from given model, even with small n DIC and LS_{CV} can be fairly strongly negatively correlated.

Above argument generalizes to any situation in which predictive distribution is approximately Gaussian (e.g., Poisson(λ) likelihood with large λ , Beta(α , β) likelihood with large ($\alpha + \beta$), etc.).

Example 3 continued. With one-sample count data (like number of hospitalizations in the *T* and *C* portions of IHGA data), people often choose between fixed- and random-effects Poisson model formulations: for i = 1, ..., n, and, e.g., with diffuse priors,

$$M_{1}: \left\{ \begin{array}{ll} \lambda & \sim & p(\lambda) \\ (y_{i}|\lambda) & \stackrel{\text{IID}}{\sim} & \text{Poisson}(\lambda) \end{array} \right\} \text{ versus } (23)$$
$$M_{2}: \left\{ \begin{array}{ll} (\beta_{0}, \sigma^{2}) & \sim & p(\beta_{0}, \sigma^{2}) \\ (y_{i}|\lambda_{i}) & \stackrel{\text{indep}}{\sim} & \text{Poisson}(\lambda_{i}) \\ (y_{i}|\lambda_{i}) & \stackrel{\text{indep}}{\sim} & \text{Poisson}(\lambda_{i}) \\ \log(\lambda_{i}) & = & \beta_{0} + e_{i} \\ e_{i} & \stackrel{\text{IID}}{\sim} & N(0, \sigma^{2}) \end{array} \right\}$$
(24)

 M_1 is special case of M_2 with $(\sigma^2 = 0, \lambda = e^{\beta_0})$; likelihood in M_2 is Lognormal mixture of Poissons (often similar to fitting negative binomial distribution, which is Gamma mixture of Poissons).

$LS_{CV} \leftrightarrow DIC$? (continued)

We conducted **partial-factorial simulation study** with factors {n = 18, 32, 42, 56, 100}, { $\beta_0 = 0.0, 1.0, 2.0$ }, { $\sigma^2 = 0.0, 0.5, 1.0, 1.5, 2.0$ } in which (**data-generating mechanism**, **assumed model**) = { $(M_1, M_1), (M_1, M_2), (M_2, M_1), (M_2, M_2)$ }; in each cell of this grid we used 100 simulation replications.



When assumed model is M_1 (fixed-effects Poisson), LS_{CV} and DIC are almost perfectly negatively correlated (we have mathematical explanation of this).



When assumed model is M_2 (random-effects Poisson), LS_{CV} and DIC are less strongly negatively correlated (DIC can misbehave with mixture models; see below), but correlation increases with n.

Example 3

As example of **correspondence between** LS_{CV} **and** DIC in real problem, IHGA data were as follows:

Distribution of number of hospitalizations in IHGA study over two-year period:

	Number of Hospitalizations										
Group	0	1	2	3	4	5	6	7	n	Mean	SD
Control	138	77	46	12	8	4	0	2	287	0.944	1.24
Treatment	147	83	37	13	3	1	1	0	285	0.768	1.01

Evidently IHGA lowered mean hospitalization rate (for these elderly Danish people, at least) by (0.944 - 0.768) =0.176, which is about $100\left(\frac{0.768 - 0.944}{0.944}\right) =$ 19% reduction from control level, a difference that's large in clinical terms.

Four **possible models** for these data (not all of them good):

- Two-independent-sample Gaussian (diffuse priors);
- One-sample Poisson (diffuse prior), pretending treatment and control λ s are equal;
- Two-independent-sample Poisson (diffuse priors), which is equivalent to fixed-effects Poisson regression (FEPR); and
- Random-effects Poisson regression (REPR), because C and T variance-to-mean ratios (VTMRs) are 1.63 and 1.32, respectively:

$$\begin{array}{ll} (y_i | \lambda_i) & \stackrel{\text{indep}}{\sim} & \text{Poisson}(\lambda_i) \\ \log(\lambda_i) & = & \beta_0 + \beta_1 x_i + e_i \\ e_i & \stackrel{\text{IID}}{\sim} & N(0, \sigma_e^2) \\ (\beta_0, \beta_1, \sigma_e^2) & \sim & \text{diffuse} \end{array}$$
(25)

where $x_i = 1$ is a **binary indicator** for T/C status.

DIC Example



To use the **DIC feature** in WinBUGS to produce the screen shot above, I fit the REPR model as usual,

did a **burn-in** of 1,000, **selected** DIC as a pull-down option from the Inference menu, **clicked** the set button in the DIC Tool window that popped up, **changed** the 1,000 to 10,000 in the updates window of the Update Tool, **clicked** update, and then **clicked** DIC in the DIC Tool when the monitoring run of 10,000 was finished—the DIC **results window** appears, with the Dbar $(\overline{D(\theta)})$, Dhat $(D(\overline{\theta}))$, pD (\hat{p}_D) , and DIC (DIC(y)) values.

DIC Example (continued)

_					
Model	$\overline{D(\theta)}$	$D(ar{ heta})$	\widehat{p}_D	DIC(y)	LS(y)
1 (Gaussian)	1749.6	1745.6	3.99	1753.5	-1.552
2 (Poisson, common λ)	1499.9	1498.8	1.02	1500.9	-1.316
3 (FEPR, different λ s)	1495.4	1493.4	1.98	1497.4	-1.314
4 (REPR)	1275.7 1274.7 1274.4	1132.0 1131.3 1130.2	143.2 143.5 144.2	1418.3 1418.2 1418.6	-1.180

DIC and **LS** results on these four models:

(3 REPR rows were based on **different monitoring runs**, all of length 10,000, to give idea of Monte Carlo noise level.)

As $\sigma_e \rightarrow 0$ in **REPR** model, you get **FEPR** model, with $p_D = 2$ parameters; as $\sigma_e \rightarrow \infty$, in effect all subjects in study have their own λ and p_D would be 572; in between at $\sigma_e \doteq 0.675$ (posterior mean), WinBUGS estimates that there are about 143 effective parameters in REPR model, but its deviance $D(\bar{\theta})$ is so much lower that it wins DIC contest hands down.



Correlation between LS and DIC across these four models is **-0.98**.

But DIC Can Misbehave

y = (0, 0, 1, 1, 1, 1, 2, 2, 2, 2, 3, 3, 3, 4, 4, 5, 6) is a data set generated from the **negative binomial** distribution with parameters (p, r) = (0.82, 10.8) (in WinBUGS notation); y has mean 2.35 and VTMR 1.22.

Using standard diffuse priors for p and r as in the BUGS examples manuals, the effective number of parameters p_D for the negative binomial model (which fits the data quite well) is estimated at -66.2:



The basic problem here is that the MCMC estimate of p_D can be **quite poor** if the marginal posteriors for one or more parameters (using the **parameterization** that defines the **deviance**) are **far from normal**; **reparameterization** helps but can still lead to **poor estimates** of p_D .

Fast (Direct) Approximation to LS_{CV}

We've seen above that DIC can sometimes provide an accurate and **fast (indirect) approximation** to LS_{CV} ; what about a **fast direct approximation**?

An obvious thing to try is the following **full-sample** version of LS: in the one-sample situation, for instance, compute a **single predictive distribution** $p(\cdot|y, M_i)$ for a future data value with each model M_i under consideration, based on the **entire data set** y (without omitting any observations), and define

$$LS_{FS}(M_i|y) = \frac{1}{n} \sum_{j=1}^{n} \log p(y_j|y, M_i).$$
 (26)

The **naive** approach to calculating LS_{CV} , when MCMC is needed to compute the predictive distributions, requires nMCMC runs, **one for each omitted observation**; by contrast LS_{FS} needs only a **single** MCMC run, making its computational speed (a) n **times faster** than naive implementations of LS_{CV} and (b) **equivalent** to that of *DIC*.

• The log score approach works equally well with parametric and nonparametric Bayesian models; *DIC* is only defined for parametric models.

• When **parametric** model M_i with parameter vector θ_i is fit via **MCMC**, the **predictive ordinate** $p(y^*|y, M_i)$ in LS_{FS} is easy to approximate: with m identically distributed (not necessarily independent) MCMC **monitoring** draws $(\theta_i)_k^*$ from $p(\theta_i|y, M_i)$,

$$p(y^*|y, M_i) = \int p(y^*|\theta_i, M_i) p(\theta_i|y, M_i) d\theta_i$$

= $E_{(\theta_i|y, M_i)} [p(y^*|\theta_i, M_i)]$ (27)
 $\doteq \frac{1}{m} \sum_{k=1}^m p(y^*|(\theta_i)_k^*, M_i).$

Example of LS_{FS} Calculations

Example. I'd like to use LS_{FS} and DIC to **compare** the **Gaussian** and t models we discussed earlier for the **NB10** data.

The files NB10-model-2.txt, NB10-data.txt, and NB10-initial-values-2.txt on the course web page contain the WinBUGS implementation of

 $M_{2}: \mu \sim N(0, \text{precision} = 1.0\text{E-6}), \sigma \sim U(0, 9.0), \\ \nu \sim U(2.0, 12.0), (y_{i}|\mu, \sigma, \nu) \stackrel{\text{IID}}{\sim} t_{\nu}(\mu, \sigma^{2})$

🗟 nb10-model3.txt 💶 🗆 🗙	anb10data
{ mu~dnorm(0.0, 1.0E-6)	list(y = c(409. 400., 406., 399., 402., 406., 401., 403., 401., 403., 398., 403., 407., 402., 401., 399., 400., 401., 405., 402., 408., 399., 399., 402., 399., 407., 401., 399., 401., 403., 400.,
sigma ~ dunif(0.0, 7.0) nu ~ dunif(2.0, 12.0)	410., 401., 407., 423., 406., 406., 402., 405., 405., 409., 399., 402., 407., 406., 413., 409., 404., 402., 404., 406., 407., 405., update100000 refree 10000
for (i in 1:n) {	411., 410., 410., 410., 401., 402., 404., 405., 392., 407., 406., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 403., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 404., 405., 408., 404., 407., 412., 406., 409., 400., 408., 404., 404., 404., 405., 404., 407., 412., 406., 409., 400., 408., 404., 404., 404., 405., 404., 407., 412., 406., 409., 400., 408., 404., 405., 406., 409., 406., 409., 406., 409., 408., 404., 406., 409., 407., 412., 406., 409., 408., 404., 404., 406., 408., 404., 407., 408., 404., 408., 404., 406., 408., 408., 404., 406., 408., 408., 404., 406., 408., 408., 408., 408., 408., 404., 408.,
y[i] ~ dt(mu, tau, nu)	401, 404, 408, 406, 408, 406, 401, 412, 393, 437, 418, 415, 408, 401, 401, 401, 401, 401, 412, 375, 409, 406, 398, 406, 406, 401, 401, 401, 401, 401, 401, 402, 412, 375, 409, 406, 398, 406, 400, 400, 400, 400, 400, 400, 400
3	403., 404.), IT = 100) Sample Monitor Tool Image: Sample Monitor Tool Image: Sample Monitor Tool
tau <- 1.0 / (sigma * sigma)	B nb10-inits3 ■□× ber 1 epc 10000 thin 1 med
y.new ~ dt(mu, tau, nu)	list(mu = 404.59, sigma = 3.0, nu = 5.0)
	stats coda uantile bgr diad auto co 95
1002 403.1 1003 404.5 1004 404.0 1005 404.8 1006 404.3 1007 404.3	nb10-model3-sigma.txt X 1001 4.851 Image: Constraint of the second
1008 404.4 1009 404.2 1010 404.4 1011 404.6 1012 404.8 1013 404.4 1014 404.4 1015 404.8	1005 4.627 Dbar Dhat pD DIC 1006 4.68 y 619.383 620.529 -1.146 618.238 1007 4.75 total 619.383 620.529 -1.146 618.238 1009 4.179 1010 4.041 1011 3.853 Image: state

I collect **100,000 monitoring iterations** for M_2 , remembering to hit the set button on the DIC tool before the monitoring begins; I use the coda button to store the μ, σ , and ν columns of the MCMC data set in files called nb10-model-2-mu.txt, nb10-model-2-sigma.txt, and nb10-model-2-nu.txt, respectively; and I hit the DIC button on the DIC tool to record that the *DIC* value for this model is **618.2** (note that *DIC* has **misbehaved** again: p_D is estimated to be -**1.1**).

I go through a **similar process** with the files NB10-model-1.txt, NB10-data.txt, and NB10-initial-values-1.txt to fit

$$\begin{split} M_1 &: \mu \sim N(0, \text{precision} = 1.0\text{E-6}), \sigma \sim U(0, 9.0), \\ & (y_i | \mu, \sigma) \stackrel{\text{IID}}{\sim} N(\mu, \sigma^2) \end{split}$$

and store the μ and σ columns of the MCMC data set in files called nb10-model-1-mu.txt and nb10-model-1-sigma.txt, respectively; this time the *DIC* value is **660.1** and DIC is **better-behaved** (p_D is estimated to be **1.9**, which is **about right**).

On the basis of *DIC* I would conclude that M_2 (**618.2** with 3 parameters) is (substantially) better than M_1 (**660.1** with 2).

Here is some R code (also available on the web page) to compute the log score values for both models.

```
> y <- dget( "nb10-data.txt" )
> y <- sort( y$y )
> mu.G <- matrix( scan( "nb10-model-1-mu.txt" ),
    100000, 2, byrow = T )[, 2]
> sigma.G <- matrix( scan( "nb10-model-1-sigma.txt" ),
    100000, 2, byrow = T )[, 2]
> mu.t <- matrix( scan( "nb10-model-2-mu.txt" ),
    100000, 2, byrow = T )[, 2]
> sigma.t <- matrix( scan( "nb10-model-2-sigma.txt" ),
    100000, 2, byrow = T )[, 2]</pre>
```

```
> nu.t <- matrix( scan( "nb10-model-2-nu.txt" ),
100000, 2, byrow = T )[ , 2 ]
```

```
> dt.s <- function( y, mu, sigma, nu ) {</pre>
    exp(lgamma((nu + 1) / 2) - ((nu + 1) / 2) *
>
      log(1 + (y - mu)<sup>2</sup> / (nu * sigma<sup>2</sup>)) -
>
      lgamma( nu / 2 ) - log( nu * pi ) / 2 - log( sigma ) )
>
> }
> LS.contributions <- matrix(0, 100, 2)
> for ( j in 1:100 ) {
    LS.contributions[ j, 1 ] <- log( mean( dt.s( y[ j ],</pre>
>
      mu.t, sigma.t, nu.t ) ) )
>
   LS.contributions[ j, 2 ] <- log( mean( dnorm( y[ j ],
>
      mu.G, sigma.G ) ) )
>
> }
> cbind( y, LS.contributions,
> 0 + LS.contributions[, 1] > LS.contributions[, 2])
                                 t
                                 better
                                 than
                       Gaussian G
               t
  [1,] 375 -8.586208 -12.204954 1
  [2,] 392 -5.349809 -4.639139 0
  [3,] 393 -5.077313 -4.362693 0
  [4,] 397 -3.903555 -3.475233 0
  [5,] 398 -3.602015 -3.309458 0
  [6,] 398 -3.602015 -3.309458 0
  [7,] 399 -3.307381 -3.166624 0
  [8,] 399 -3.307381 -3.166624 0
```

[9,]	399	-3.307381	-3.166624	0
[10,]	399	-3.307381	-3.166624	0
[11,]	399	-3.307381	-3.166624	0
[12,]	399	-3.307381	-3.166624	0
[13,]	399	-3.307381	-3.166624	0
[14,]	400	-3.028685	-3.046933	1
[15,]	400	-3.028685	-3.046933	1
[16,]	400	-3.028685	-3.046933	1
[17,]	400	-3.028685	-3.046933	1
[18,]	401	-2.778176	-2.950552	1
[19,]	401	-2.778176	-2.950552	1
[20,]	401	-2.778176	-2.950552	1
[21,]	401	-2.778176	-2.950552	1
[22,]	401	-2.778176	-2.950552	1
[23,]	401	-2.778176	-2.950552	1
[24,]	401	-2.778176	-2.950552	1
[25,]	401	-2.778176	-2.950552	1
[26,]	401	-2.778176	-2.950552	1
[27,]	401	-2.778176	-2.950552	1
[28,]	401	-2.778176	-2.950552	1
[29,]	401	-2.778176	-2.950552	1
[30,]	402	-2.571441	-2.877618	1
[31,]	402	-2.571441	-2.877618	1
[32,]	402	-2.571441	-2.877618	1
[33,]	402	-2.571441	-2.877618	1
[34,]	402	-2.571441	-2.877618	1
[35,]	402	-2.571441	-2.877618	1
[36,]	402	-2.571441	-2.877618	1
[37,]	402	-2.571441	-2.877618	1
[38,]	403	-2.426129	-2.828236	1
[39,]	403	-2.426129	-2.828236	1
[40,]	403	-2.426129	-2.828236	1
[41,]	403	-2.426129	-2.828236	1
[42,]	403	-2.426129	-2.828236	1
[43,]	403	-2.426129	-2.828236	1
[44,]	404	-2.358212	-2.802475	1

[45,]	404	-2.358212	-2.802475	1
[46,]	404	-2.358212	-2.802475	1
[47,]	404	-2.358212	-2.802475	1
[48,]	404	-2.358212	-2.802475	1
[49,]	404	-2.358212	-2.802475	1
[50,]	404	-2.358212	-2.802475	1
[51,]	404	-2.358212	-2.802475	1
[52,]	404	-2.358212	-2.802475	1
[53,]	405	-2.376305	-2.800373	1
[54,]	405	-2.376305	-2.800373	1
[55,]	405	-2.376305	-2.800373	1
[56,]	405	-2.376305	-2.800373	1
[57,]	405	-2.376305	-2.800373	1
[58,]	406	-2.477698	-2.821932	1
[59,]	406	-2.477698	-2.821932	1
[60,]	406	-2.477698	-2.821932	1
[61,]	406	-2.477698	-2.821932	1
[62,]	406	-2.477698	-2.821932	1
[63,]	406	-2.477698	-2.821932	1
[64,]	406	-2.477698	-2.821932	1
[65,]	406	-2.477698	-2.821932	1
[66,]	406	-2.477698	-2.821932	1
[67,]	406	-2.477698	-2.821932	1
[68,]	406	-2.477698	-2.821932	1
[69,]	406	-2.477698	-2.821932	1
[70,]	407	-2.649778	-2.867123	1
[71,]	407	-2.649778	-2.867123	1
[72,]	407	-2.649778	-2.867123	1
[73,]	407	-2.649778	-2.867123	1
[74,]	407	-2.649778	-2.867123	1
[75,]	407	-2.649778	-2.867123	1
[76,]	407	-2.649778	-2.867123	1
[77,]	407	-2.649778	-2.867123	1
[78,]	408	-2.875393	-2.935880	1
[79,]	408	-2.875393	-2.935880	1
[80,]	408	-2.875393	-2.935880	1

[81,]	408	-2.875393	-2.935880	1	
[82,]	408	-2.875393	-2.935880	1	
[83,]	409	-3.137771	-3.028107	0	
[84,]	409	-3.137771	-3.028107	0	
[85,]	409	-3.137771	-3.028107	0	
[86,]	409	-3.137771	-3.028107	0	
[87,]	409	-3.137771	-3.028107	0	
[88,]	410	-3.422943	-3.143672	0	
[89,]	410	-3.422943	-3.143672	0	
[90,]	410	-3.422943	-3.143672	0	
[91,]	410	-3.422943	-3.143672	0	
[92,]	411	-3.720225	-3.282413	0	
[93,]	412	-4.021816	-3.444136	0	
[94,]	412	-4.021816	-3.444136	0	
[95,]	412	-4.021816	-3.444136	0	
[96,]	413	-4.322196	-3.628616	0	
[97,]	415	-4.905384	-4.064801	0	
[98,]	418	-5.710652	-4.882504	0	
[99,]	423	-6.845648	-6.656119	0	
[100,]	437	-9.016222	-13.896384	1	
> sum(> ler [1] 0.7	LS.c ngth(71	contributio	ms[,1]>	> LS.contributions[,	2])/
# Thus # 71% c	t mo of th	odel is pre ne data poi	dictively k nts.	petter than Gaussian f	or
LS.t <-	- mea	an(LS.cont	ributions[, 1])	
LS.G <-	- mea	an(LS.cont	ributions[,2])	
c(LS.t	t, LS	5.G)			
[1] -3	.0823	331 -3.2621	42		

Although it's not immediately **obvious**, the **log score** for the t model (-3.08) is **substantially higher** than that for the Gaussian model (-3.26), so LS and DIC have reached the **same conclusion** here.

```
> plot( y, LS.contributions[, 1],
> ylim = c( min( LS.contributions ),
> max( LS.contributions ) ),
> ylab = 'Log Score Contributions' )
> lines( y, LS.contributions[, 1], lty = 1 )
> points( y, LS.contributions[, 2], pch = 2 )
> lines( y, LS.contributions[, 2], lty = 2 )
> legend( 397.5, -10, c( "t", "Gaussian" ), pch = c( 1, 2 ) )
```



The *t* model **fits better** both **in the tails** (where the **most influential observations** are from the Gaussian point of view) and in the **center** (where **most** of the data values are).

Asymptotic Properties of LS_{FS}

Recall the claim that LS_{CV} approximates expectation of logarithmic utility:

$$E[U(M_i|y)] \approx LS_{CV} = \frac{1}{n} \sum_{j=1}^{n} \log p(y_j|M_i, y_{-j})$$
 (28)

Berger et al. (2005) recently proved that difference between LHS and RHS of (28) does not vanish for large n but is instead $O_p(\sqrt{n})$.

(However **unpleasant**, this fact does not automatically invalidate use of LS_{CV} as estimated expected utility, since when comparing two models we effectively look at the **difference** between two LS_{CV} values, and the discrepancy should largely **cancel out**.)

We have proved for a simple model that LS_{FS} is free from this deficiency: the difference between $E[U(M_i|y)]$ and $LS_{FS} = \frac{1}{n} \sum_{j=1}^{n} \log p(y_j|y, M_i)$ is $O_p(1)$ (we expect the general proof to go through as well).

Q: Does this asymptotic superiority of LS_{FS} over LS_{CV} translate into better small-sample performance?

LS_{CV} , LS_{FS} and DICModel Discrimination

We now have three behavioral rules: maximize LS_{CV} , maximize LS_{FS} , minimize DIC.

With (e.g.) two models to choose between, how accurately do these behavioral rules discriminate between M_1 and M_2 ?

Example: Recall that in **earlier simulation study**, for i = 1, ..., n, and with **diffuse priors**, we considered

$$M_{1}: \left\{ \begin{array}{ll} \lambda & \sim & p(\lambda) \\ (y_{i}|\lambda) & \stackrel{\text{IID}}{\sim} & \text{Poisson}(\lambda) \end{array} \right\} \text{ versus}$$
$$M_{2}: \left\{ \begin{array}{ll} (\beta_{0}, \sigma^{2}) & \sim & p(\beta_{0}, \sigma^{2}) \\ (y_{i}|\lambda_{i}) & \stackrel{\text{indep}}{\sim} & \text{Poisson}(\lambda_{i}) \\ \log(\lambda_{i}) & = & \beta_{0} + e_{i} \\ e_{i} & \stackrel{\text{IID}}{\sim} & N(0, \sigma^{2}) \end{array} \right\}$$

Model Discrimination (continued)

As extension of previous simulation study, we generated data from M_2 and computed LS_{CV} , LS_{FS} , and DIC for models M_1 and M_2 in full-factorial grid $\{n = 32, 42, 56, 100\}$, $\{\beta_0 = 0.0, 1.0\}$, $\sigma^2 = 0.1, 0.25, 0.5, 1.0, 1.5, 2.0\}$, with 100 simulation replications in each cell, and monitored percentages of correct model choice (here M_2 is always correct).

Examples of **results** for (e.g.) LS_{CV} :

% Correct Decision eta_0			Mea	Mean Absolute Difference in LS_{CV} eta_0			
σ^2	0	1	σ^2	0	1		
0.10	31	47	0.10	0.001	0.002		
0.25	49	85	0.25	5 0.002	0.013		
0.50	76	95	0.50	0.017	0.221		
1.00	97	100	1.00	0.237	4.07		
1.50	98	100	1.50) 1.44	17.4		
2.00	100	100	2.00) 12.8	63.9		

n = 32

Even with *n* only **32**, LS_{CV} makes the right model choice **more than 90% of the time** when $\sigma^2 > 0.5$ for $\beta_0 = 1$ and when $\sigma^2 > 1.0$ for $\beta_0 = 0$.

Model Discrimination (continued)



The plots above compare **Bayesian decision-theoretic** power curves for LS_{CV} (solid lines), LS_{FS} (long dotted lines), and *DIC* (short dotted lines) (column 1: $\beta_0 = 0$; column 2: $\beta_0 = 1$).

Remarkably, not only is LS_{FS} much quicker computationally than LS_{CV} , it's also more accurate at identifying the correct model than LS_{CV} or DIC.

To summarize, in computational efficiency

$$LS_{CV} < DIC \doteq LS_{FS} \tag{29}$$

and in **fixed-** and **random-effects Poisson modeling** the results in **model discrimination power** are

$$LS_{CV} \doteq DIC < LS_{FS} \tag{30}$$

Why Not Bayes Factors?

Much has been written about use of **Bayes factors** for model choice (e.g., Jeffreys 1939, Kass and Raftery 1995; excellent recent book by O'Hagan and Forster 2004 devotes almost **40 pages** to this topic).

Why not use **probability scale** to choose between M_1 and M_2 ?

$$\begin{bmatrix} \underline{p(M_1|y)}\\ \overline{p(M_2|y)} \end{bmatrix} = \begin{bmatrix} \underline{p(M_1)}\\ \overline{p(M_2)} \end{bmatrix} \cdot \begin{bmatrix} \underline{p(y|M_1)}\\ \overline{p(y|M_2)} \end{bmatrix}$$
(31)
$$\begin{pmatrix} \text{posterior}\\ \text{odds} \end{pmatrix} = \begin{pmatrix} \text{prior}\\ \text{odds} \end{pmatrix} \cdot \begin{pmatrix} \text{Bayes}\\ \text{factor} \end{pmatrix}$$

Kass and Raftery (1995) note that

$$\log \left[\frac{p(y|M_1)}{p(y|M_2)} \right] = \log p(y|M_1) - \log p(y|M_2)$$
(32)
= $LS^*(M_1|y) - LS^*(M_2|y),$
where

$$LS^{*}(M_{i}|y) \equiv \log p(y|M_{i})$$

= $\log [p(y_{1}|M_{i}) p(y_{2}|y_{1}, M_{i}) \cdots p(y_{n}|y_{1}, \dots, y_{n-1}, M_{i})]$
= $\log p(y_{1}|M) + \sum_{j=2}^{n} \log p(y_{j}|y_{1}, \dots, y_{j-1}, M_{i}).$

Thus log Bayes factor equals difference between models in something that looks like a log score, i.e., aren't LS_{CV} and LS_{FS} equivalent to choosing M_i whenever the Bayes factor in favor of M_i exceeds 1?

$LS \neq BF$

No; crucially, LS* is defined via sequential prediction of y_2 from y_1 , y_3 from (y_1, y_2) , etc., whereas LS_{CV} and LS_{FS} are based on averaging over all possible out-of-sample predictions.

This distinction really matters: as is well known, with diffuse priors Bayes factors are hideously sensitive to particular form in which diffuseness is specified, but this defect is entirely absent from LS_{CV} and LS_{FS} (and from other properly-defined utility-based model choice criteria).

Example: Integer-valued data $y = (y_1, \ldots, y_n)$;

 $M_1 = \text{Geometric}(\theta_1)$ likelihood with $\text{Beta}(\alpha_1, \beta_1)$ prior on θ_1 ;

 $M_2 = \text{Poisson}(\theta_2)$ likelihood with $\text{Gamma}(\alpha_2, \beta_2)$ prior on θ_2 .

Bayes factor in favor of M_1 over M_2 is

$$\frac{\Gamma(\alpha_1+\beta_1)\Gamma(n+\alpha_1)\Gamma(n\bar{y}+\beta_1)\Gamma(\alpha_2)(n+\beta_2)^{n\bar{y}+\alpha_2}\left(\prod_{i=1}^n y_i!\right)}{\Gamma(\alpha_1)\Gamma(\beta_1)\Gamma(n+n\bar{y}+\alpha_1+\beta_1)\Gamma(n\bar{y}+\alpha_2)\beta_2^{\alpha_2}}.$$

Diffuse priors: take $(\alpha_1, \beta_1) = (1, 1)$ and $(\alpha_2, \beta_2) = (\epsilon, \epsilon)$ for some $\epsilon > 0$.

Bayes factor reduces to

$$\frac{\Gamma(n+1)\Gamma(n\bar{y}+1)\Gamma(\epsilon)(n+\epsilon)^{n\bar{y}+\epsilon}\left(\prod_{i=1}^{n}y_{i}!\right)}{\Gamma(n+n\bar{y}+2)\Gamma(n\bar{y}+\epsilon)\epsilon^{\epsilon}}.$$

$LS \neq BF$ (continued)

This goes to $+\infty$ as $\epsilon \downarrow 0$, i.e., you can make the evidence in favor of the Geometric model over the Poisson as large as you want as a function of a quantity near 0 that scientifically you have no basis to specify.

By contrast, e.g.,

$$LS_{CV}(M_1|y) = \log\left[\frac{(\alpha_1 + n - 1)\Gamma(\beta_1 + s)}{\Gamma(\alpha_1 + n + \beta_1 + s)}\right] + \frac{1}{n}\sum_{i=1}^n \log\left[\frac{\Gamma(\alpha_1 + n - 1 + \beta_1 + s_i)}{\Gamma(\beta_1 + s_i)}\right]$$

and

$$LS_{CV} = (M_2|y) = \frac{1}{n} \sum_{i=1}^{n} \log \left[\frac{\Gamma(\alpha_2 + s)}{\Gamma(y_i + 1)\Gamma(\alpha_2 + s_i)} \\ \cdot \left(\frac{\beta_2 + n}{\beta_2 + n + 1} \right)^{\alpha_2 + s_i} \left(\frac{1}{\beta_2 + n + 1} \right)^{y_i} \right]$$

(with similar expressions for LS_{FS}); both of these quantities are **entirely stable** as a function of (α_1, β_1) and (α_2, β_2) near zero.

(Various **attempts** have been made to **fix** this defect of Bayes factors, e.g., {partial, intrinsic, fractional} Bayes factors, well calibrated priors, conventional priors, intrinsic priors, expected posterior priors, ... (e.g., Pericchi 2004); all of these methods appear to require an appeal to **ad-hockery** which is **not required by the log score approach**.)

(Some **bridges** can be built between **LS** and **BF**, e.g., Berger et al. (2005) re-interpret LS_{CV} as the "Gelfand-Dey (1994) **predictive Bayes factor**" BF^{GD} ; connections like these are the subject of **ongoing investigation**.)

What LS_{FS} Is Not

(1) Likelihood part of (parametric) model

 $M_j: (y_i|\theta_j, M_j) \stackrel{\text{IID}}{\sim} p(y_i|\theta_j, M_j) (j = 1, 2), \text{ with prior } p(\theta_j|M_j) \text{ for model } M_j.$

Ordinary Bayes factor involves comparing quantities of the form

$$p(y|M_j) = \int \left[\prod_{i=1}^n p(y_i|\theta_j, M_j)\right] p(\theta_j|M_j) d\theta_j,$$

= $E_{(\theta_j|M_j)} L(\theta_j|y, M_j),$ (33)

i.e., Bayes factor involves comparing **expectations** of **likelihoods** with respect to the **priors** in the models under comparison (this is **why ordinary Bayes factors behave so badly with diffuse priors**).

Aitkin (1991; **posterior Bayes factors**): compute expectations instead with respect to the **posteriors**, i.e.,

PBF: favor model M_1 if $\log \overline{L}_1^A > \log \overline{L}_2^A$, where

$$\log \bar{L}_j^A = \log \int \left[\prod_{i=1}^n p(y_i|\theta_j, M_j)\right] p(\theta_j|y, M_j) \, d\theta_j.$$
(34)

This **solves** the problem of sensitivity to a diffuse prior but **creates new problems of its own**, e.g., it's **incoherent**.

It may **seem** at first glance (e.g., O'Hagan and Forster (2004) think so) that **PBF** is the same thing as LS_{FS} : favor model M_1 if

$$n LS_{FS}(M_1|y) > n LS_{FS}(M_2|y).$$
 (35)

But not so:

$$nLS_{FS}(M_j|y) = \log \prod_{i=1}^n \left[\int p(y_i|\theta_j, M_j) \, p(\theta_j|y, M_j) \, d\theta_j \right], \quad (36)$$

and this is **not the same** because the **integral** and **product** operators **do not commute**.

What LS_{FS} Is Not (continued)

Also, some people (e.g., Geweke (2005)) like to compare models based on the **posterior expectation of the log likelihood** (this is **one of the ingredients** in *DIC*), and this is **not the same** as LS_{FS} either: by **Jensen's inequality**

$$nLS_{FS}(M_j|y) = \sum_{i=1}^{n} \log p(y_i|y, M_j)$$

$$= \sum_{i=1}^{n} \log \int p(y_i|\theta_j, M_j) p(\theta_j|y, M_j) d\theta_j$$

$$= \sum_{i=1}^{n} \log E_{(\theta_j|y, M_j)} L(\theta_j|y_i, M_j)$$

$$> \sum_{i=1}^{n} E_{(\theta_j|y, M_j)} \log L(\theta_j|y_i, M_j) \quad (37)$$

$$= E_{(\theta_j|y, M_j)} \sum_{i=1}^{n} \log L(\theta_j|y_i, M_j)$$

$$= E_{(\theta_j|y, M_j)} \log \prod_{i=1}^{n} L(\theta_j|y_i, M_j)$$

$$= E_{(\theta_j|y, M_j)} \log L(\theta_j|y, M_j).$$

When Is a Model Good Enough?

 LS_{FS} method described here (not LS* method) can stably and reliably help in choosing between M_1 and M_2 ; but suppose M_1 has a (substantially) higher LS_{FS} than M_2 .

This doesn't say that M_1 is **adequate**—it just says that M_1 is better than M_2 , i.e., what about model specification question (2): Is M_1 good enough?

As mentioned above, a **full judgment of adequacy** requires **real-world input** (to what purpose will the model be put?), but you can answer a somewhat related question—**could the data have arisen from a given model**?—in a general way by **simulating** from that model many times, **developing** a distribution of (e.g.) LS_{FS} values, and **seeing how unusual** the actual data set's log score is in this distribution (Draper and Krnjajić 2004).

This is related to the **posterior predictive model-checking** method of Gelman, Meng and Stern (1996); however, this sort of thing cannot be done **naively**, or result will be **poor calibration**—indeed, Robins et al. (2000) demonstrated that the Gelman et al. procedure may be (sharply) **conservative**.

Using modification of idea in Robins et al., we have developed method for accurately calibrating the log score scale.

Inputs to our procedure: (1) A **data set** (e.g., with regression structure); (2) A **model** (can be parametric, non-parametric, or semi-parametric).

Simple example: data set y = (1, 2, 2, 3, 3, 3, 4, 6, 7, 11), n = 10.

Given model (*)

- $(\lambda) \sim \text{Gamma}(0.001, 0.001)$ (38)
- $(y_i|\lambda) \stackrel{\mathrm{IID}}{\sim} \mathsf{Poisson}(\lambda)$

Calibrating LS_{FS} Scale

Step 1:

Calculate LS_{FS} for this data set; say get $LS_{FS} = -1.1$; call this **actual log score** (ALS).

Obtain posterior for λ given y based on this data set; call this **actual posterior**.

Step 2:

for (i in 1:m1) {

make a lambda draw from the actual posterior; call it lambda[i]

generate a data set of size n from the second line of model (*) above, using lambda = lambda[i]

compute the log score for this generated
 data set; call it LS[i]

}

Output of this loop is a vector of log scores; call this V.LS.

Locate ALS in distribution of LS_{FS} values by computing percentage of LS_{FS} values in V.LS that are \leq ALS; call this percentage **unadjusted actual tail area** (say this is 0.22).

So far this is just Gelman et al. with LS_{FS} as the **discrepancy function**.

We know from our own simulations and the literature (Robins et al. 2000) that this tail area (a *p*-value for a **composite null hypothesis**, e.g., Poisson(λ) with λ unspecified) is **conservative**, i.e., with the 0.22 example above an adjusted version of it that is well calibrated would be **smaller**.

Calibrating LSFS Scale (continued)

We've **modified** and implemented one of the ways suggested by Robins et al., and we've shown that it does indeed work even in rather small-sample situations, although our approach to implementing the basic idea can be **computationally intensive**.

Step 3:

for (j in 1:m2){

- make a lambda draw from the actual posterior; call it lambda*.
- generate a data set of size n from the second line
 of model (*) above, using lambda = lambda*;
 call this the simulated data set

repeat steps 1, 2 above on this simulated data set

}

The result will be a vector of unadjusted tail areas; call this **V.P**.

Compute the percentage of tail areas in V.P that are \leq the unadjusted actual tail area; this is the **adjusted actual tail area**.

Calibrating LS_{FS} Scale (continued)

The claim is that the 3-step procedure above is well-calibrated, i.e., if the sampling part of model (*) really did generate the observed data, the distribution of adjusted actual tail areas obtained in this way would be uniform, apart from simulation noise.

```
Step 3 in this procedure solves the calibration problem by applying the old idea that if X \sim F_X then F_X(X) \sim U(0, 1).
```

This claim can be verified by building a **big loop** around steps 1–3 as follows:

Choose a lambda value of interest; call it lambda.sim

for (k in 1:m3) {

generate a data set of size n from the second line of model (*) above, using lambda = lambda.sim; call this the validation data set

repeat steps 1-3 on the validation data set

}

The result will be a vector of **adjusted P-values**; call this **V.Pa**.

We have **verified** (via simulation) in several simple (and some less simple) situations that the values in V.Pa are close to U(0, 1) in distribution.

Two **examples**—Poisson(λ) and Gaussian(μ, σ^2):

Uncalibrated p-values

Null Poisson model: Uncalibrated p-values





Uncalibrated p-values

Null Gaussian model: Uncalibrated p-values



Calibrated p-values

Null Gaussian model: Calibrated p-values vs uniform(0,1)



R Implementation

Here's some R code (available at the course web site) to implement our method for calibrating the log score scale in a one-sample Poisson setting, applied first to the length of stay data from part 2b and then to a simulated data set that was not generated by the Poisson model.

```
> print( y <- c( 0, 1, 1, 1, 1, 1, 2, 2, 2, 2, 3, 3, 4, 6 ) )</pre>
 [1] 0 1 1 1 1 1 2 2 2 2 3 3 4 6
> print( epsilon <- 0.001 )</pre>
[1] 0.001
> ln.poisson.gamma <- function( y, alpha, beta ) {</pre>
+
    lgamma( alpha + y ) + alpha * log( beta /
+
      (beta + 1)) + y * log(1 / (beta + 1)) -
+
      lgamma( alpha ) - lgamma( y + 1 )
+
+
+ }
> step1 <- function( y, epsilon ) {</pre>
+
    n <- length( y )</pre>
+
+
    s <- sum( y )
+
+
    als <- mean( ln.poisson.gamma( y, epsilon + s,
+
      epsilon + n ) )
+
+
    return( c( n, s, als ) )
+
+
+ }
> print( step1.result <- step1( y, epsilon ) )</pre>
[1] 14.00000 29.00000 -1.71309
```

So the **actual log score** for the LoS data set is -1.71, but is this **unusually small if the data really were Poisson**?

```
> step2 <- function( n, s, epsilon, als, m1 ) {</pre>
+
    lambda <- rgamma( m1, epsilon + s, epsilon + n )</pre>
+
+
    ls <- rep( 0, m1 )
+
+
    for ( i in 1:m1 ) {
+
+
      y.star <- rpois( n, lambda[ i ] )</pre>
+
+
      s.star <- sum( y.star )</pre>
+
+
      ls[ i ] <- mean( ln.poisson.gamma( y.star,</pre>
+
        epsilon + s.star, epsilon + n ) )
+
+
    }
+
+
    uata <- sum( ls <= als ) / m1
+
+
    write( ls, "ls.out" )
+
+
    return( uata )
+
+
+ }
> m1 <- 1000
>
> print( step2.result <- step2( step1.result[ 1 ],</pre>
    step1.result[ 2 ], epsilon, step1.result[ 3 ], m1 ) )
+
[1] 0.418
> v.ls <- scan( "ls.out" )
Read 1000 items
>
> hist( v.ls, nclass = 20, probability = T,
    main = '', xlab = 'uncalibrated log score' )
+
>
> abline( v = step1.result[ 3 ] )
```



The actual log score doesn't look at all unusual in this plot, but recall from the discussion above that it may not yet be properly calibrated.

```
> step3 <- function( y, epsilon, m1, m2 ) {</pre>
+
    step1.result <- step1( y, epsilon )</pre>
+
+
    n <- step1.result[ 1 ]</pre>
+
+
    s.actual <- step1.result[ 2 ]</pre>
+
+
    uata <- step2( step1.result[ 1 ], step1.result[ 2 ],</pre>
+
     epsilon, step1.result[ 3 ], m1 )
+
+
    v.p <- rep( 0, m2 )
+
```

```
for ( j in 1:m2 ) {
+
+
       lambda.star <- rgamma( 1, epsilon + s.actual,</pre>
+
        epsilon + n )
+
+
      y.sim <- rpois( n, lambda.star )</pre>
+
+
      step1.result <- step1( y.sim, epsilon )</pre>
+
+
      v.p[ j ] <- step2( step1.result[ 1 ],</pre>
+
         step1.result[ 2 ], epsilon, step1.result[ 3 ], m1 )
+
+
    }
+
+
    aata <- sum( v.p <= uata ) / m2</pre>
+
+
    write( v.p, "v.p.out" )
+
+
    return( aata )
+
+
+ }
> m2 <- 100
>
> print( step3.result <- step3( y, epsilon, m1, m2 ) )</pre>
[1] 0.4
```

Here the **recalibration** has **not had much effect**, but (as the plots above showed) **this will not always be the case**.

```
> v.p <- scan( "v.p.out" )
Read 100 items
>
> hist( v.p, nclass = 20, probability = T, xlim = c( 0, 1 ),
+ main = '', xlab = 'calibrated tail areas' )
> abline( v = step2.result )
```



For a **second example** let's look at a **data set** generated as a **lognormal mixture of Poissons** with a **substantial VTMR**.

```
> n <- 10
>
> e <- rnorm( n, 0.0, 0.5 )
>
mu <- 0
>
lambda <- rep( 0, n )</pre>
```

```
> y <- rep( 0, n )
> for ( i in 1:n ) {
+
    lambda[ i ] <- exp( mu + e[ i ] )</pre>
+
+
    y[i] <- rpois( 1, lambda[ i ] )</pre>
+
+
+ }
> print( y <- sort( y ) )</pre>
[1] 0 0 0 1 1 1 2 3 4 4
> var( y ) / mean( y )
[1] 1.555556
> print( step1.result <- step1( y, epsilon ) )</pre>
[1] 10.000000 16.000000 -1.715601
> print( step2.result <- step2( step1.result[ 1 ],</pre>
    step1.result[ 2 ], epsilon, step1.result[ 3 ], m1 ) )
+
[1] 0.178
> v.ls <- scan( "ls.out" )
> hist( v.ls, nclass = 20, probability = T,
    main = '', xlab = 'uncalibrated log score' )
+
> abline( v = step1.result[ 3 ] )
```



> m2 <- 1000

```
> print( step3.result <- step3( y, epsilon, m1, m2 ) )</pre>
```

[1] 0.099

```
So here's an example where the uncalibrated tail area is about twice as big as it should be.
```

```
> v.p <- scan( "v.p.out" )
> hist( v.p, nclass = 20, probability = T, xlim = c( 0, 1 ),
+ main = '', xlab = 'calibrated tail areas' )
> abline( v = step2.result )
```



The true calibrated tail-area distribution is far from uniform, so 0.178 is actually substantially farther out in the true tail than it seems.

Conclusions

 {Exchangeability judgments plus nonparametric (BNP) modeling} = Bayesian model specification in many problems.

• BNP is one way to avoid the dilemma posed by Cromwell's Rule in Bayesian model specification; three-way cross-validation (3CV) is another.

 Model choice is really a decision problem and should be approached via MEU, with a utility structure that's sensitive to the real-world context.

 When the goal is to make an accurate scientific summary of what's known about something, the predictive log score has a sound generic utility basis and can yield stable and accurate model specification decisions.

• *DIC* can be thought of as a fast approximation to the **leave-one-out predictive log score** (LS_{CV}) , but *DIC* can behave **unstably** as a function of **parameterization**.

• The full-sample log score (LS_{FS}) is *n* times faster than naive implementations of LS_{CV} , has better small-sample model discrimination power than either LS_{CV} or DIC, and has better asymptotic behavior than LS_{CV} .

• Generic Bayes factors are highly unstable when context suggests diffuse prior information; many methods for fixing this have been proposed, most of which seem to require an appeal to ad-hockery which is absent from the LS_{FS} approach.

• The basic Gelman et al. (1996) method of posterior predictive model-checking is **badly calibrated**: when it gives you a tail area of, e.g., **0.4**, the calibrated equivalent may well be **0.04** or even **0.004**.

• We have modified an **approach** suggested by Robins et al. (2000) to help answer the question "Could the data have arisen from model *M*?" in a **well-calibrated** way.

References

Bernardo JM, Smith AFM (1994). Bayesian Theory. New York: Wiley.

- Dey D, Mueller P, Sinha D (1998). *Practical Nonparametric and Semiparametric Bayesian Statistics*. New York: Springer Verlag (Lecture Notes in Statistics, Volume 133).
- de Finetti B (1930). Funzione caratteristica de un fenomeno aleatorio. *Mem. Acad. Naz. Lincei*, **4**, 86–133.
- de Finetti B (1937). La prévision: ses lois logiques, ses sources subjectives. Ann. Inst. H. Poincaré, **7**, 1–68.
- de Finetti B (1938). Sur la condition d'equivalence partielle. Actualités Scientifiques et Industrielles, **739**.
- de Finetti B (1990). *Theory of Probability*. New York: Wiley Classics Library.
- Draper D (1995). Assessment and propagation of model uncertainty (with discussion). *Journal of the Royal Statistical Society Series B*, **57**, 45–97.
- Draper D (1996). Utility, sensitivity analysis, and cross-validation in Bayesian model-checking. *Statistica Sinica*, **6**, 760–767 (discussion of "Posterior predictive assessment of model fitness via realized discrepancies," by A Gelman, X-L Meng, and H Stern).
- Draper D, Fouskakis D (2000). A case study of stochastic optimization in health policy: problem formulation and preliminary results. *Journal of Global Optimization*, **18**, 399–416.
- Draper D, Fouskakis D (2004). Stochastic optimization methods for costeffective quality assessment in health. Submitted.
- Draper D, Krnjajić M (2005). Three-way cross-validation for well-calibrated model exploration. In preparation.
- Draper D, Hodges J, Mallows C, Pregibon D (1993). Exchangeability and data analysis (with discussion). *Journal of the Royal Statistical Society Series A*, **156**, 9–37.
- Fouskakis D, Draper D (2002). Stochastic optimization: a review. *International Statistical Review*, **70**, 315–349.

References (continued)

- Geisser S, Eddy WF (1979). A predictive approach to model selection. Journal of the American Statistical Association, **74**, 153–160.
- Gelfand AE, Dey DK, Chang H (1992). Model determination using predictive distributions, with implementation via sampling-based methods (with discussion). In *Bayesian Statistics 4* (Bernardo JM, Berger JO, Dawid AP, Smith AFM, editors), Oxford: Oxford University Press, 147–167.
- Gelman A, Meng X-L, Stern H (1996). Posterior predictive assessment of model fitness via realized discrepancies (with discussion). *Statistica Sinica*, 6, 733–760.
- Good IJ (1950). Probability and the Weighing of Evidence. London: Griffin.
- Hendriksen C, Lund E, Stromgard E (1984). Consequences of assessment and intervention among elderly people: a three year randomised controlled trial. *British Medical Journal*, **289**, 1522–1524.
- Jeffreys H (1939). Theory of Probability. Oxford: Oxford University Press.
- Kass RE, Raftery AE (1995). Bayes factors. *Journal of the American Statistical Association*, **90**, 773–795.
- Key JT, Pericchi LR, Smith AFM (1998). Bayesian model choice: what and why? (with discussion). In *Bayesian Statistics 6*, Bernardo JM, Berger JO, Dawid AP, Smith AFM (editors). Oxford University Press, 343–370.
- O'Hagan A, Forster J (2004). *Bayesian Inference*, second edition. London: Arnold.
- Pericchi L (2004). Model selection and hypothesis testing based on objective probabilities and Bayes factors. Manuscript.
- Robins JM, van der Vaart A, Ventura V (2000). Asymptotic distribution of *P* values in composite null models. *Journal of the American Statistical Association*, **95**, 1143–1156.
- Spiegelhalter DJ, Best NG, Carlin BR, van der Linde A (2002). Bayesian measures of model complexity and fit (with discussion). *Journal of the Royal Statistical Society Series B*, **64**, 583–616.
- Walker S, Damien P, Lenk P (2004). On priors with a Kullback-Leibler property. *Journal of the American Statistical Association*, **99**, 404–408.